

Greenhouse gas emission from the soils fertilized with liquid pig slurry (LPS) in Tifton 85 bermudagrass pasture in tropical savanna

Adilson Amorim Brandão^{1*}, Eduardo Guimarães Couto², Renato de Aragão Ribeiro Rodrigues³, Oscarlina Lúcia Santos Weber², Osvaldo Borges Pinto Júnior⁴

¹Federal Institute of Mato Grosso do Sul, Naviraí, 79950-000, Mato Grosso do Sul, Brazil

²Federal University of Mato Grosso, Department of Soils and Rural Engineering, Cuiabá, 78060-900, Mato Grosso, Brazil

³Embrapa Soils, Rio de Janeiro, 22460-000, Rio de Janeiro, Brazil

⁴University of Cuiabá, Cuiabá, 78065-900, Mato Grosso, Brazil

*Corresponding author: aabrandaojr@gmail.com

Abstract

Soils have important roles in the global budgets of the greenhouse gases. The liquid pig slurry (LPS) in pastures has high potential as a fertilizer but could have a direct influence on emission of greenhouse gasses. This study evaluated the effects of the application of LPS and inorganic mineral fertilization during the rainy and dry seasons on the emissions of CO₂, CH₄ and N₂O in pastures planted with Tifton-85. The following treatments were tested: Control - no fertilization; LPS30 - 30 m³ ha⁻¹; LPS60 - 60 m³ ha⁻¹; LPS90 - 90 m³ ha⁻¹ and inorganic mineral fertilization. Gasses were sampled using static chambers first during the months of March and April, then in June and July. Fertilization with LPS caused an increase in the flux of CO₂ and CH₄ during the first hours after its application, and CO₂ emissions are greater during the rainy than in the dry season. However, the application of LPS in Tifton-85 pasture during rainy periods did not show high potential for emission of CO₂, in contrast to application during the dry season. Fertilization with LPS increases the emission of N₂O, and this varies as a function of the volume of LPS applied and the experimental conditions. The application of LPS in Tifton-85 pasture has a high potential for N₂O emission during the rainy season, but the magnitude is similar to that resulting from inorganic mineral fertilization.

Keywords: swine wastewater, soil gas fluxes, carbon dioxide, methane, nitrous oxide.

Abbreviations: LPS_liquid pig slurry; DAA_day after application; GHG_greenhouse gas; IC_intensive collection.

Introduction

The expansion of pasture for livestock production is the main responsible for land use change (LUC) emissions contributing for 6% of global emissions from livestock. The LUC emission from soybean is also a substantial driver of LUC emissions, covering 3.2% of global emissions from livestock (Gerber et al., 2013). The gas emission in Brazil is mostly caused by cattle, expansion of pasture for production of feed ingredients such as soybean (Karstensen et al., 2013) for pigs and poultry. In fact, pigs and poultry are mostly fed ingredients containing grain and soy products (Kebreab et al., 2016). Brazilian production of pork is increasing, with a 28.2% growth rate expected by the end of 2029 (MAPA, 2019). However, a consequence of pig farming is the generation and concentration of a large volume of excrement, which represents an environmental risk and requires measures to be taken to reduce negative impacts on the surrounding environment (Ceretta et al., 2005). Pork farming in a confined system generates concentrated liquid slurry in specific areas on rural properties (Damasceno, 2010). For example, in the state of Mato Grosso, the estimated annual production of slurry in 2018 by

pig herd amounted ~ 2.94 million, according IBGE (2018). It corresponds to a daily production of 7 liters of liquid slurry per animal (Oliveira, 2004), by which the annual slurry production is approximately 7.51 million m³.

The application of liquid pig slurry (LPS) in pastures is a viable alternative for its use because as a function of its chemical characteristics it has great potential to be used as a fertilizer (Seidel et al., 2010). Therefore, LPS could partially or totally substitute inorganic mineral fertilizer and make a significant contribution to crop production and reduce input costs (Scherer et al., 2012). The most common form of use of LPS is in application to the surface of agricultural soils due to its lower operational costs and ease of application in most cultivation systems (Friederichs et al., 2019), and also because it favors the accumulation of nutrients in the root zone of agricultural soils (Veiga et al., 2012; Friederichs, 2019). However, the inadequate use of LPS can increase the risk of microbiological contamination of groundwater, accumulation of toxic elements, nutrient imbalances and soil impermeabilization (Seganfredo, 2000), besides having a direct influence on the emissions of carbon dioxide (CO₂),

methane (CH₄) and nitrous oxide (N₂O), and on volatilization of ammonia (NH₃) (Carvalho and Hentz, 2014).

The emission of CO₂ from the soil occurs as a function of respiration of roots and organisms together with decomposition of organic residuals (Carvalho et al., 2010). These processes are influenced by the application of LPS to the soil due to an increase in organic carbon, and consequently an increase in microbial respiration and activity (Webb et al., 2010). Furthermore, fertilization with LPS contributes to the development of agricultural crops by increasing plant root systems and the input of plant residual materials to the soil.

In research conducted in a controlled laboratory environment, CO₂ flux was 125 mg C m⁻² h⁻¹ one day after application (DAA) of LPS (200 kg N ha⁻¹) in an uncultivated clay soil, while in a soil without application of LPS the flux was 40 mg C m⁻² h⁻¹ (Jarecki et al., 2008). In a no-till soil cultivated with a wheat and corn rotation of CO₂ flux rates varied between 60 and 208 mg m⁻² h⁻¹ and after treatment of LPS at rate of 140 kg N ha⁻¹. These rates remained higher than those of the control treatment during the first 18 days after application (Grave et al., 2015).

The flux of soil CH₄ is the result of production of this gas in the soil by methanogenic bacteria under anaerobic conditions, and also the oxidation that occurs through the action of methanotrophic bacteria in the presence of oxygen (Le Mer and Roger, 2001). The oxidation of methane is generally greater than its production in well-drained and aerated soils. However, the application of different forms of nitrogen can considerably reduce the rates of oxidation of CH₄ (Suwanwaree and Robertson, 2005).

In a study conducted in New Zealand, CH₄ flux was measured during 90 days after the application of 60 m³ ha⁻¹ of LPS to a pasture. The flux of soil CH₄ was 3,933 µg C m⁻² h⁻¹ immediately after application of LPS and continued at a lower rate until the 7th DAA. From the 8th DAA onward the measured fluxes indicated absorption of atmospheric methane by the soil (Sherlock et al., 2002).

In Frederico Westphalen-RS, Brazil, treatments applying LPS were evaluated with the objective of measuring greenhouse gas emissions as a function of the method of application to no-till soil cultivated with a wheat and corn rotation (Arenhardt, 2016). The results showed that the accumulated CH₄ emissions from soil where LPS was applied on the surface were negative, meaning that methane was absorbed, although there was soil CH₄ efflux soon after LPS application.

The volatilization of ammonia (NH₃) is considered to be the principal mechanism of loss of nitrogen (N) after application of liquid residuals to the soil (Smith et al., 2009). This reduces the potential of nitrogen as a fertilizer and contributes to environmental contamination (Rochette et al., 2008). Losses of ammonia are generally high and can reach 75% of NH₄⁺-N when liquid residuals, principally those from pigs, are applied with incorporation into the soil (Thompson and Meisinger, 2002). Although ammonia is not a greenhouse gas (GHG) itself, but its emission to the atmosphere and subsequent deposition on the soil can result in production of nitrous oxide (N₂O) as a by-product of the microbial nitrification process, and also as a product of microbial denitrification (Zaman and Blennerhassett, 2010). The Inter-governmental Panel on Climate Change (IPCC) estimates that, on average, 1.25% of total N from fertilizer or manure applied to soil is emitted as N₂O (IPCC, 2007).

In a study conducted in the Brazilian state of Rio Grande do Sul, the flux of soil N₂O in a no-till system using an oat mulch was increased by 160 µg N m⁻² h⁻¹ four hours after the application of 40 m³ ha⁻¹, compared to the treatment without LPS. The accumulated emission of N₂O 28 days after application was 93.8 g N ha⁻¹ (Giacomini et al., 2006). This dynamic of more intense N₂O flux during the first DAA after LPS application has been reported in the literature for temperate climate conditions (Rochette et al., 2008; Pelster et al., 2012) as well as for subtropical (Aita et al., 2014, Aita et al., 2015).

The potential of LPS for the use as a substitute for inorganic mineral fertilization in agricultural production, including for forage grasses in pasture systems, is widely reported in the literature (Drumond et al., 2006; Medeiros et al., 2007; Camargo et al., 2011; Silva et al., 2015). However, it is necessary to evaluate the dimensions of the impacts of LPS application with respect to the emissions of CO₂, CH₄ and N₂O in pasture areas, especially in regions with a tropical climate. These GHG are important for atmospheric chemistry and its radiation balance as a function of their lifetimes in the atmosphere, and also because of the values for relative global warming potential (GWP) of CH₄ and N₂O which are respectively about 25 and 298 times greater than that of CO₂ (IPCC, 2007).

The objective of this study was to evaluate the effects of the application of LPS and inorganic mineral fertilization during the rainy and dry seasons on the emissions of CO₂, CH₄ and N₂O in pastures planted with Tifton-85.

Results and Discussion

Daily average flux of CO₂, CH₄ and N₂O during the intensive collection campaigns

Before application of fertilizer two intensive collection (IC) campaigns were conducted. The CO₂ fluxes were similar between the two treatments. Starting on the first day after fertilizer application (DAA), in the first IC as well as for the second IC the flux was increased in the treatments LPS30, LPS60 and LPS90 (Figs 1A and 1B). In the first IC, treatment LPS90 had a CO₂ flux above 500 mg m⁻² h⁻¹ and was statistically different from the other treatments (F_{4,12}= 3.506; p<0.05). In the second IC, treatments LPS30, LPS60 and LPS90 had similar CO₂ flux values, but differed from those for the Control and FertMin treatments (F_{4,12}= 4.941; p<0.05). In general, the greater the rate of application of LPS, the higher the emission values for CO₂ on the 1st DAA. The CO₂ flux was decreased starting from 2nd DAA until the end of the evaluation period for both the first and second ICs, with no differences between treatments.

The application of LPS to the soil stimulates an increase in CO₂ emission during the first hours after application, with about 59% of the organic carbon added in LPS being emitted as CO₂, independent of the application method to the soil (Giacomini et al., 2007). The increase in CO₂ emission during the first hours after LPS application is reported in the literature (Aita, Port and Giacomini, 2006; Giacomini et al., 2007; Giacomini and Aita, 2008; Arenhardt, 2016) and could be linked to the liberation of CO₂ dissolved in the LPS, as well as CO₂ formed from HCO₃⁻ and CO₃²⁻ generated during storage of this material (Carvalho and Hentz, 2014). Evidence for this shown by the fact that the CO₂ flux values on the days preceding the application of the LPS to the soil

varied between 1 and 2 kg ha⁻¹ h⁻¹ independent of the treatment.

During the 1st IC, several rainfall events apparently influenced the CO₂ flux in all treatments, with peaks of C-CO₂ flux on March 27 and 29, and April 1 (Fig 1A). This effect is due to the fact that precipitation increases soil biological activity, and this is reflected in greater respiration and CO₂ emission rates (Luo and Zhou, 2006a, 2006b).

The fluxes of CH₄ were equal before the application of the treatments for each IC, showing emission peaks after the application of the LPS. From the 7th DAA, the fluxes diminished and became similar between treatments (Figs 1C and 1D). In the 1st IC there were differences between treatments only on day 1 and from the 5th to the 7th DAA (Fig 1C). On the 1st DAA, treatment LPS90 had an emission of 642.7 µg m⁻² h⁻¹ and was different from the other treatments (F_{4,12}= 6.869; p<0.05). The FertMin treatment had a C-CH₄ flux of -7.4 µg m⁻² h⁻¹ on the 5th DAA and was different from the other treatments (F_{4,12}= 6.533; p<0.01), which all showed a positive flux for methane. On the 6th and 7th DAA, the treatment LPS30 had higher values than the other treatments, with a flux of 20.1 µg m⁻² h⁻¹ and 38.0 µg m⁻² h⁻¹, respectively.

The flux values for CH₄ during the 2nd IC was increased in the treatments with LPS a few hours after its application (Fig 1D), as also shown for the 1st IC. On the 1st DAA, CH₄ emissions for LPS60 and LPS90 were 45.7 and 47.0 µg m⁻² h⁻¹, respectively, and were different from the other treatments (F_{4,12}= 4.028; p<0.05). Treatments LPS90 and FertMin had similar or greater values than those from the other treatments on the 5th DAA (F_{4,12}= 5.178; p<0.05).

The emission peaks of methane are most likely related to volatilization of dissolved CH₄ soon after application of LPS (Sherlock et al., 2002). Damasceno (2010) reported results that showed that methane emission increases in the soil with addition of residuals from pork farming, with a methane emission rate of 600 µg m⁻² h⁻¹ soon after application of LPS on the soil surface. However, in the current study the increase in CH₄ flux between the 5th and 7th DAA during the 1st IC could be related to rainfall events that saturated the soil pores and formed anaerobic micro-environments that are favorable for methanogenesis.

In each IC, the majority of treatments showed negative values for CH₄ flux (influx or absorption), indicating consumption of methane. This corroborated with Piva et al. (2013) in cultivated pasture on an Oxisol in Paraná. An influx of methane is expected for an aerated and well-structured soil such as the one in the current study because this soil type is considered to be a methane sink (Saggar et al., 2008). Before the fertilizer application, N₂O fluxes were similar between the treatments in both ICs (Figs 1E and 1F). After the application of the LPS, from the 1st to the 3rd DAA, treatment LPS90 had greater emissions than the other treatments (Fig 1E). Between the 4th and the 6th DAA, an N-N₂O emission in the inorganic mineral fertilization treatment was increased, which were similar to emissions from LPS90 on the 5th DAA and greater than those from the other treatments between the 4th and the 6th DAA. Starting on the 7th DAA, average flux of N₂O was lower and there was a difference between treatments only between the 7th and 9th DAA, with greater flux in LPS90 and FertMin (Fig 1E).

In the 2nd IC, there were differences between treatments only for the 1st DAA (Fig 1F), with LPS60 and LPS90 having N₂O fluxes greater than the other treatments. In general,

during this period, the emission of N₂O in the LPS treatments was greater.

The average N₂O flux values in this study were lower than those reported by Denega (2009) principally after application of LPS, where this author observed emissions above 600 µg m⁻² h⁻¹ up to 48 hours after LPS application on soil covered with a corn mulch. However, the results from the current study are within the range reported in the literature (Giacomini et al., 2006).

The increase in N₂O emission soon after LPS application to the soil has been reported in a large range of studies (Giacomini et al., 2006; Rochette et al., 2008; Denega, 2009; Arenhardt, 2016). The increase in N₂O flux due to LPS application shown during the first two periods in the current study could be attributed to three factors: 1) the organic carbon in the LPS is used by aerobic microorganisms in the soil as a carbon and energy source. This increases their activity and demand for O₂, which could create anaerobic sites. Certain types of microbes can respire using nitrate present in the soil as the final electron receptor in the respiratory chain reducing it to N₂O 2) the rapid nitrification of NH₃-N applied with the LPS could result in the production of N₂O by certain bacteria that possess the nitrite reductase enzyme 3) the liquid fraction could facilitate the occurrence of environments that are deficient in O₂ depending on soil humidity and on the dose of LPS added to the soil, where nitrate could be denitrified and emitted to the atmosphere in the form of N₂O (Giacomini et al., 2006).

The increase in the average daily fluxes in the treatment FertMin was observed in the 1st IC in agreement with the literature (Aita et al., 2015; Arenhardt, 2016). The presence of mineral N combined with the humid soil creates a favorable environment for the denitrification process (Zanatta et al., 2010), which could explain the increase in the N₂O flux observed in this study.

The lower emissions in the 2nd IC compared to the 1st IC could be related to low soil humidity during June 22 to July 5. The elevated capacity for water infiltration into the soil may favor the greater concentration of O₂ in the soil profile, which represents an unfavorable situation for the denitrification process and consequently emission of N₂O (Jantalia et al., 2008).

Accumulated emission of CO₂, CH₄ and N₂O during the intensive collection campaigns

In the 1st IC, the accumulated emissions of CO₂ did not show any differences between the treatments (Fig 2A), indicating that during a period with more rainfall the application of LPS does not cause a significant increase in the flux of CO₂. This could be explained by conditions that favor mineralization of carbon present in the soil including good levels of soil humidity and higher temperatures. However, during the 2nd IC, the application of LPS resulted in an increase in accumulated emission, with emissions from LPS90 being 3-fold greater compared to those of the Control (Fig 2A). Although the values of the accumulated emissions of CO₂ of the 2nd IC were inferior to those of the 1st IC, the use of LPS for fertilization has high potential emissions of this gas during periods with lower rainfall.

The accumulated emissions of CH₄ in the LPS treatments were greater in each IC (Fig 2B). During the 1st IC, LPS90 had greater accumulated emission of methane than the other treatments, and together with LPS30 and LPS60 were the

Table 1. Chemical composition of liquid pig slurry for each application date.

Month/year	N	P	K	Ca	Mg	S	Zn	Cu	Mn	Fe
	g L ⁻¹						mg L ⁻¹			
March/2015	2.10	0.70	0.40	0.15	0.09	0.41	1.20	2.10	0.60	2.60
June/2015	1.10	0.07	0.73	0.18	0.16	0.02	1.80	1.20	0.40	3.60

N=nitrogen; P=phosphorus; K=potassium; Ca=calcium; Mg=magnesium; S=sulfur; Zn=zinc; Cu=copper; Mn=manganes; and Fe=iron.

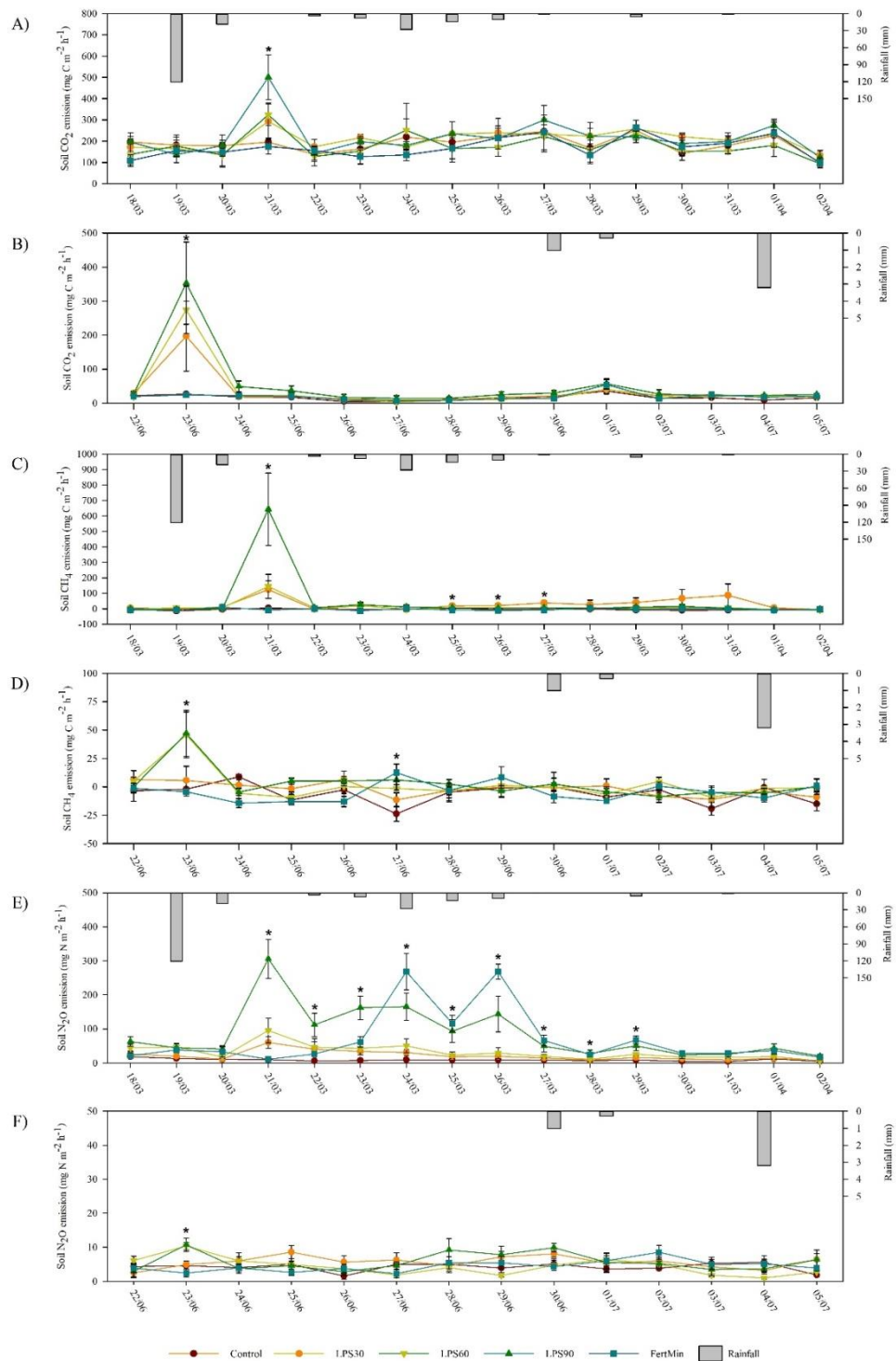


Fig 1. Average daily flux for CO_2 in the 1st IC (A) and the 2nd IC (B); CH_4 in the 1st IC (C) and the 2nd IC (D); and N_2O in the 1st IC (E) and the 2nd IC (F). Vertical bars represent the standard error of the mean ($n=4$). *significant difference by the Tukey's test ($p<0.05$).

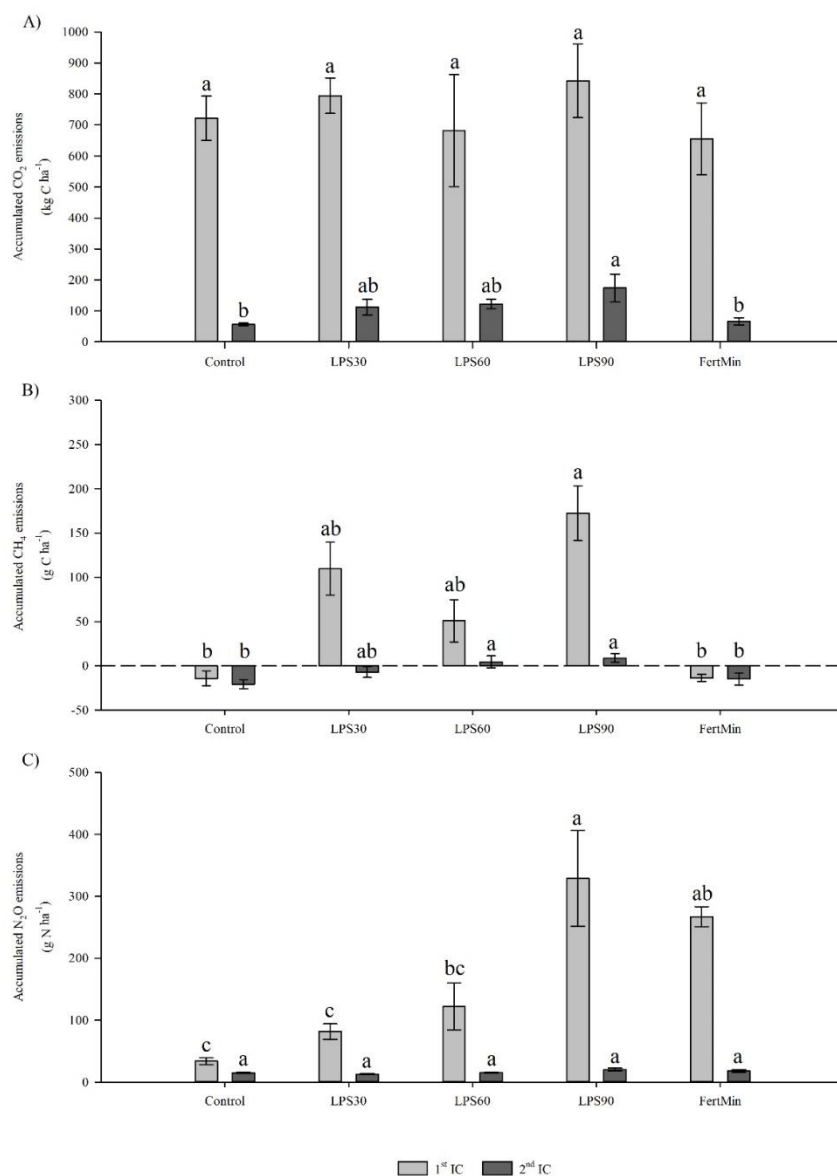


Fig 2. Accumulated emission of: (a) CO₂, (b) CH₄ and (c) N₂O in the 1st and 2nd IC. The vertical bars represent the standard error of the mean (n=4). Means followed by the same lowercase letters in to each intensive collection campaign does not differ statistically by the Tukey's test (p<0.05).

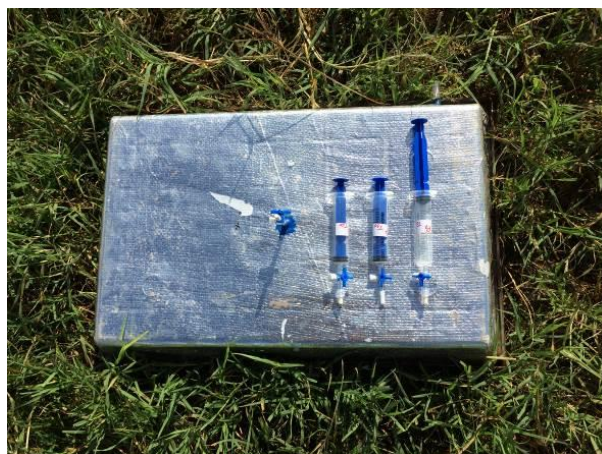


Fig 3. PVC chamber covered with a thermal insulation layer and three gas samplers (syringes) used for each collection period.

sources of this gas. In the 2nd IC, LPS60 and LPS90 had greater accumulated emissions of CH₄ and was a source of this gas to the atmosphere. In each of the intensive collection periods, the Control and FertMin treatments had negative accumulated emissions (influx), which characterized them as sinks of methane during the evaluation period (Fig 2B).

The accumulated emission of N₂O during the 1st IC showed a significant difference between treatments, with LPS90 and FertMin having greater emissions than the other treatments (Fig 2C). Greater accumulated emissions in the treatments with LPS and inorganic mineral fertilizer, but with no statistical differences, were reported by other studies (Arenhardt, 2016). In the 2nd IC, the accumulated emissions of N₂O did not differ between treatments (Fig 2C). This fact could be associated with losses of N through volatilization of NH₃ when LPS was applied to the soil surface, together with low soil humidity and a high level of oxygenation, which could have limited denitrification during this evaluation period (Aita et al., 2014).

Materials and Methods

Study area and description of experiment

The experiment was conducted under field conditions in the experimental area of the Rio Verde Foundation for Research and Technological Development, located at 13° 00' 02" S, 55° 58' 15" W and at an altitude of 387 m, in the municipality of Lucas do Rio Verde – MT, on a Red-Yellow dystrophic Oxisol of clay texture. The chemical analysis of the soil from 0 to 0.20 m layer presented pH=4.53 (H₂O); 14.22 mg dm⁻³ P; 106.28 mg dm⁻³ K; 2.27 cmolc dm⁻³ Ca; 0.93 cmolc dm⁻³ Mg; 15.15 cmolc dm⁻³ H + Al; 0.38 cmolc dm⁻³ Al and 17.94% base saturation (V%). The climate is Aw and tropical with dry winter season according to the Köppen classification system. The experimental design was randomized blocks with five treatments and four repetitions, for a total of 20 plots with an area of 55 m² each, cultivated since March 2014 with a Tifton 85 bermudagrass cultivar (*Cynodon dactylon*). The following treatments were tested: Control – no fertilization; LPS30 - 30 m³ ha⁻¹ of LPS; LPS60 - 60 m³ ha⁻¹ of LPS; LPS90 - 90 m³ ha⁻¹ of LPS, and FertMin - inorganic mineral fertilization. Treatment FertMin-inorganic mineral fertilization was applied in January 2015 at a rate of 200 kg N in the form of urea, 70 kg P₂O₅ as single superphosphate, and 400 kg K₂O as potassium chloride, all on a per hectare basis. The applications of the treatments in the experimental area occurred monthly between January and June 2015. However, the application of LPS was done on March 20, 2015 and June 23, 2015, always at the end of the afternoon, and the nutrient concentrations of the LPS applied on these dates are shown in Table 1.

Sample collection

Sample collection was conducted using *in situ* closed-cover static chambers (Fig 3) based on methodology adapted from the literature (Nogueira et al., 2015).

The samples were collected during the morning between 8-10 a.m. (GMT-4) a period considered representative of average daily GHG flux (Zuchello, 2010). Immediately after coupling the chamber on the metallic base, a sample of atmospheric air was collected using a plastic syringe with a

volume of 20 cm³. Thirty minutes after coupling of chamber, the first air sample was collected from its interior, followed by another sample at 60 minutes, for a total of three samples taken at 0, 30, and 60 minutes. To facilitate sample collection, the chamber had a connecting port on its top that is regulated using a three-way plastic valve. The syringe was connected to this valve by another three-way plastic valve, which ensured proper sealing after sample collection (Rodrigues and Mello, 2012). After each collection, the samples were transferred to flasks that had their air content removed by vacuum. Additionally, air and soil temperatures were measured during sampling.

Intensive sample collection campaigns (IC) were done in two distinct periods, with the first IC done between March 18 and April 20, 2015, and the second IC between June 22 and July 23, 2015. The IC were conducted during the three days that preceded the application of the LPS, and also on the day of the application and the period afterward, with the goal of accompanying the pattern of GHG flux resulting from the fertilization treatments.

Laboratory analyses

The determination of gas concentrations was done at Embrapa Agrossilvipastoril using a gas chromatograph (model Shimadzu GC-2014) with FID and ECD detectors and an automatic injector. The calibration curves were determined using three gas standards for CO₂, CH₄ and N₂O (White Martins). These standards were: 308,700; 714,600 and 5097,000 micromol/mol, for CO₂; 1,402; 9,722 and 100,300 micromol/mol, for CH₄; and 360,000; 832,000 and 2080,000 nanomol/mol, for N₂O. The time for which the samples were analyzed was 8 minutes.

Statistical analysis

Average and accumulated flux data for CO₂, CH₄ and N₂O were processed for descriptive analysis and analyzed for normality (Shapiro-Wilk), homogeneity of variances (Levene), analysis of variance (ANOVA), and treatment means were compared using the Tukey's test (p<0.05). When necessary, the data were *log* transformed in order to homogenize the variances and normalize the frequency distribution. These results were transformed using the inverse operation to present the data in their original units.

Conclusion

The influence of fertilization with LPS increased the fluxes of CO₂ and CH₄ from the first hours after its application. During the period with greater precipitation, CO₂ emissions were higher than in the dry season. However, the application of LPS in Tifton-85 pasture during rainy periods did not have high potential for emission of CO₂, in contrast to this application during the dry season. The soil fertilized with LPS is occasionally a source of CH₄ during periods with greater rainfall. Fertilization with LPS increases the emission of N₂O, and this varies as a function of the volume of LPS applied and the experimental conditions. The application of LPS in Tifton-85 pasture has a high potential for N₂O emission during the rainy season, but the magnitude is similar to that resulting from inorganic mineral fertilization. The emission of GHGs by the LPS treatments in this study were similar to

those observed in the area that received inorganic mineral fertilizer.

Acknowledgments

This study was financed in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – Brasil (CAPES) – Finance Code 001 and BRF S.A.

References

- Aita C, Gonzatto R, Miola ECC, Santos DB, Rochette P, Angers DA, Chantigny MH, Pujol SB, Giacomini DA, Giacomini SJ (2014) Injection of dicyandiamide-treated pig slurry reduced ammonia volatilization without enhancing soil nitrous oxide emissions from no-till corn in southern Brazil. *J Environ Qual*. 43(3): 789-800.
- Aita C, Port O, Giacomini SJ (2006) Dinâmica do nitrogênio no solo e produção de fitomassa por plantas de cobertura no outono/inverno com o uso de dejetos de suínos. *Rev Bras Cienc Solo*. 30(5): 901-910.
- Aita C, Schirrmann J, Pujol SB, Giacomini SJ, Rochette P, Angers DA, Chantigny MH, Gonzatto R, Giacomini DA, Doneda A (2015) Reducing nitrous oxide emissions from a maize-wheat sequence by decreasing soil nitrate concentration: effects of split application of pig slurry and dicyandiamide. *Eur J Soil Sci*. 66(2): 359-368.
- Arenhardt, MH (2016) Emissões de gases de efeito estufa em resposta ao modo de aplicação de dejetos de suínos e ao uso de inibidor de nitrificação na sucessão trigo/milho em latossolo. Santa Maria: Universidade Federal de Santa Maria, 2016. Dissertation, 77 f.
- Camargo SC, Mesquita EE, Castagnara DD, Neres MA, Oliveira PSR (2011) Efeito da aplicação de dejetos de suínos na concentração de minerais na parte aérea de capins Tifton 85. *Scientia Agrar Par*. 10(2): 51-62.
- Carvalho NL, Hentz, P. Impactos ambientais da fertilização orgânica em sistemas agropecuários na região sul-brasileira. *REGET*. 18(1): 340-352.
- Carvalho JLN, Avanzi JC, Silva MLN, Mello CRD, Cerri CEP (2010) Potencial de sequestro de carbono em diferentes biomas do Brasil. *Rev Bras Cienc Solo*. 34(2): 277-290.
- Ceretta CA, Basso CJ, Pavinato PS, Trentin EE, Girotto E (2005) Produtividade de grãos de milho, produção de matéria seca e acúmulo de nitrogênio, fósforo e potássio na rotação aveia preta/milho/nabo forrageiro com aplicação de dejetos líquidos de suínos. *Cienc Rural*. 35(6): 1287-1295.
- Damasceno F (2010) Injeção de dejetos líquidos de suínos no solo e inibidor de nitrificação como estratégias para reduzir as emissões de amônia e óxido nítrico. Santa Maria: Universidade Federal de Santa Maria, 2010. Dissertation, 93 f.
- Denega GL (2009) Emissão de óxido nítrico e dióxido de carbono após aplicação de dejetos de suínos e bovinos em um argissolo. Santa Maria: Universidade Federal de Santa Maria, 2010. Dissertation, 122 f.
- Drumond LCD, Zanini JR, Aguiar APA, Rodrigues GP, Fernandes ALT (2006) Produção de matéria seca em pastagem de Tifton 85 irrigada, com diferentes doses de dejetos líquidos de suíno. *Eng Agric*. 26(2): 426-433.
- Friederichs A, Borges Júnior WS, Roters DF, Mafra AL (2019) Emissão de CO₂ do solo pela aplicação de fertilizantes orgânicos e minerais em ambiente controlado. *Rev Agropec Catarin*. 32(2): 86-91.
- Gerber PJ, Steinfeld H, Henderson B, Mottet A, Opio C, Dijkman J, Faluccci A, Tempio G (2013) Tackling climate change through livestock and a global assessment of emissions and mitigation opportunities. Food and Agriculture Organization of the United Nations (FAO), Rome.
- Giacomini S, Recous S, Mary B, Aita C (2007) Simulating the effects of N availability, straw particle size and location in soil on C and N mineralization. *Plant and Soil*. 301(1): 289-301.
- Giacomini SJ, Aita C (2008) Emissão de dióxido de carbono após aplicação de dejetos líquidos e cama sobreposta de suínos. *Pesqui Agropecu Bras*. 43(1): 107-114.
- Giacomini SJ, Jantalia CP, Aita C, Urquiaga S (2006) Emissão de óxido nítrico com a aplicação de dejetos líquidos de suínos em solo sob plantio direto. *Pesqui Agropecu Bras*. 41(11): 1653-1661.
- Grave R, Nicoloso RS, Cassol PC, Aita C, Corrêa JC, Costa MD, Fritz DD (2014) Short-term carbon dioxide emission under contrasting soil disturbance levels and organic amendments. *Soil Tillage Res*. 146: 184-192.
- IBGE – Instituto Brasileiro de Geografia e Estatística. Pesquisa da Pecuária Municipal – PPM, 2018. <<https://www.ibge.gov.br/estatisticas/economicas/agricultura-e-pecuaria/9107-producao-da-pecuaria-municipal.html?=&t=resultados>> 25 Set.
- Jantalia CP, Santos HP, Urquiaga S, Boddey RM, Alves BJR (2008) Fluxes of nitrous oxide from soil under different crop rotations and tillage systems in the south of Brazil. *Nutr Cycling Agroecosyst*. 82(2): 161-173.
- Jarecki MK, Parkin TB, Chan AS, Hatfield JL, Jones R (2008) Greenhouse gas emissions from two soils receiving nitrogen fertilizer and swine manure slurry. *J Environ Qual*. 37(4): 1432-1438.
- Karstensen J, Peters GP, Andrew RM (2013) Attribution of CO₂ emissions from Brazilian deforestation to consumers between 1990 and 2010. *Environ Res Lett*. 8: 1-7.
- Kebreab E, Liedke A, Caro D, Diemling S, Binder M, Finkbeiner M (2016) Environmental impact of using specialty feed ingredients in swine and poultry production: a life cycle assessment. *J Animal Sci*. 94: 2664-2681.
- Le Mer J, Roger P (2001) Production, oxidation, emission and consumption of methane by soils: A review. *Eur J Soil Biol*. 37(1): 25-50.
- IPCC – Intergovernmental Panel on Climate Change (2007) Climate change 2007: The Physical Science Basis. contribution of working group I to the fourth assessment report of the IPCC, (eds. Solomon S, Qin D, Manning M, Chen Z, Marquis M, Averyt KB, Tignor M, Miller HL). Cambridge University Press, Cambridge.
- Luo Y, Zhou X (2006a) Processes of CO₂ production in soil. In: Luo Y, Zhou X (eds) soil respiration and the environment, Academic Press, Elsevier, San Diego, 35.
- Luo Y, Zhou X (2006b) Controlling factors. In: Luo Y, Zhou X (eds) soil respiration and the environment, Academic Press, Elsevier, San Diego, 79.
- Medeiros LT, Rezende AV, Vieira PF, Neto FRC, Valeriano AR, Casali AO, Gastaldello Junior AL (2007) Produção e qualidade da forragem de capim-marandu fertirrigada com dejetos líquidos de suínos. *Rev Bras Zootec*. 36(2): 309-318.

- MAPA – Ministério da Agricultura Pecuária e Abastecimento (2019). Projeções do Agronegócio: Brasil 2018/19 a 2028/29 projeções de longo prazo. Brasília, 126 p.
- Nogueira AKS, Rodrigues RAR, Castro BS, Nogueira TF, Silva JN, Behling M, Mombach M, Armacolo N, Silveira JG (2015) Emissões de óxido nitroso e metano do solo em áreas de recuperação de pastagens na amazônia matogrossense. *Quim Nova*. 38(7): 937-943.
- Oliveira PAV (2004) Tecnologias para o manejo de resíduos na produção de suínos: manual de boas práticas. Embrapa Suínos e Aves, 109 p.
- Pelster DE, Chantigny MH, Rochette P, Angers DA, Rieux C, Vanasse A (2012) Nitrous oxide emissions respond differently to mineral and organic nitrogen sources in contrasting soil types. *J Environ Qual*. 41(2): 427-435.
- Piva JT, Dieckow J, Bayer C, Zanatta JA, Moraes A, Tomazi M, Pauletti V, Barth G, Piccolo MC (2013) Soil gaseous N₂O and CH₄ emissions and carbon pool due to integrated crop-livestock in a subtropical Ferralsol. *Agric Ecosyst Environ*. 190: 87-93.
- Rochette P, Angers DA, Chantigny MH, MacDonald JD, Gasser MO, Bertrand N (2008) Reducing ammonia volatilization in a no-till soil by incorporating urea and pig slurry in shallow bands. *Nutri Cycling Agroecosyst*. 84(1): 71-80.
- Rodrigues RAR, Mello, WZD (2012) Fluxos de óxido nitroso em solos com cobertura de floresta ombrófila densa montana na Serra dos Órgãos, Rio de Janeiro. *Quim Nova*. 35(8) 1549-1553.
- Saggar S, Tate KR, Giltrap D, Singh JS (2008) Soil-atmosphere exchange of nitrous oxide and methane in New Zealand terrestrial ecosystems and their mitigation options: A review. *Plant Soil*. 309(1): 25-42.
- Scherer EE, Baldissera IT, Miranda M (2012) Produção e qualidade de forragem e atributos do solo influenciados por adubação orgânica e nitrogenada. *Epagri*, 52 p.
- Seganfredo MA (2000) A questão ambiental na utilização de dejetos de suínos como fertilizante do solo. Embrapa Suínos e Aves, 35 p.
- Seidel EP, Gonçalves Junior AC, Vanin JP, Strey L, Schwantes D, Nacke H (2010) Aplicação de dejetos de suínos na cultura do milho cultivado em sistema de plantio direto. *Acta Sci Technol*. 32(2): 113-117.
- Sherlock R, Sommer SG, Khan RZ, Wood CW, Guertal EA, Frenay JR, Dawson CO, Cameron KC (2002) Ammonia, Methane, and Nitrous Oxide emission from pig slurry applied to a pasture in New Zealand. *J Environ Qual*. 31(5): 1491-1501.
- Silva ADA, Lana ÂMQ, Lana RMQ, Costa AMD (2015) Fertilização com dejetos suínos: influência nas características bromatológicas da *Brachiaria decumbens* e alterações no solo. *Eng Agric*. 35(2): 254-265.
- Smith E, Gordon R, Bourque C, Campbell A, Générmont S, Rochette P, Mkhabela M (2009) Simulated management effects on ammonia emissions from field applied manure. *J Environ Manage*. 90(8): 2531-2536.
- Suwanwaree P, Robertson GP (2005) Methane oxidation in forest, successional, and no-till agricultural ecosystems. *Soil Sci Soc Am J*. 69(6): 1722-1729.
- Thompson RB, Meisinger JJ (2002) Management factors affecting ammonia volatilization from land-applied cattle slurry in the Mid-Atlantic USA. *J environ qual*. 31(4): 1329-1338.
- Veiga MD, Pandolfo CM, Balbinot Junior AA, Spagnollo E (2012) Chemical attributes of a hapludox soil after nine years of pig slurry application. *Pesq Agropec Bras*. 47(12): 1766-1773.
- Webb J, Pain B, Bittman S, Morgan J (2010) The impacts of manure application methods on emissions of ammonia, nitrous oxide and on crop response - A review. *Agr Ecosyst Environ*. 137(1): 39-46.
- Zaman M, Blennerhassett J (2010) Effects of the different rates of urease and nitrification inhibitors on gaseous emissions of ammonia and nitrous oxide, nitrate leaching and pasture production from urine patches in an intensive grazed pasture system. *Agr Ecosyst Environ*. 136(3):236-246.
- Zanatta JA, Bayer C, Vieira FCB, Gomes J, Tomazi M (2010) Nitrous oxide and methane fluxes in south Brazilian gleysol as affected by nitrogen fertilizers. *Rev Bras Cienc Solo*. 34(5): 1653-1665.
- Zuchello F (2010) Emissões de óxido nitroso (N₂O) de cambissolo cultivado com cana-de-açúcar em Campos dos Goytacazes: impacto de adubações com vinhaça e uréia. Seropédica: Universidade Federal Rural do Rio de Janeiro, 2010. Dissertation, 45 f.